

Integrated Flux Balance Analysis (iFBA)

Metabolische Netzwerke SS 15

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Integrated Flux Balance Analysis (iFBA)

Covert/Xiao/Chen/Karr'08

"A major advantage of rFBA - requiring few kinetic parameters - could be a weakness in situations where the kinetic parameters have been determined and capture information not contained in rFBA."

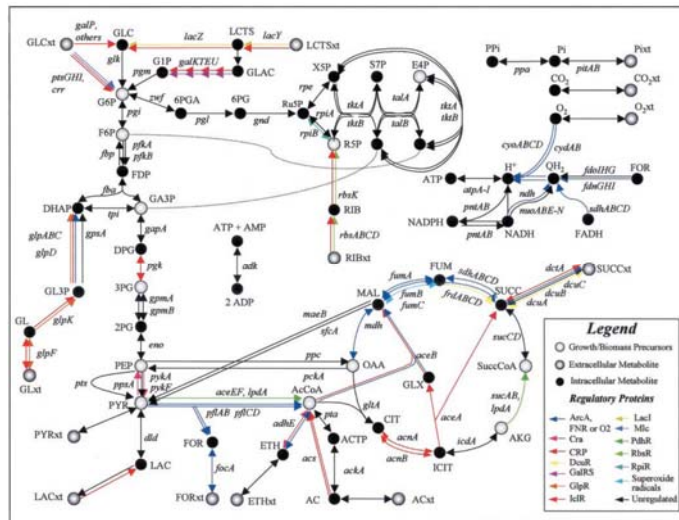
- Integrate available kinetic information into rFBA models (iFBA).
- More precisely: combine rFBA and ODE model by identifying values to pass from either model to the other.

Software available:

<http://covertlab.stanford.edu/projects/iFBA/>

rFBA model of central metabolism in E.coli

Covert/
Palsson'02



ODE model of E.Coli carbohydrate uptake

Kremling et al.07

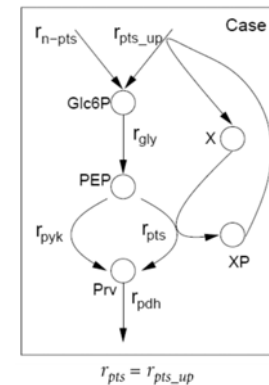
Differential equations

$$\dot{G6P} = r_{n-pts} + r_{pts_up} - r_{gly}$$

$$\dot{PEP} = 2r_{gly} - r_{pyk} - r_{pts}$$

$$\dot{Prv} = r_{pts} + r_{pyk} - r_{pdh}$$

$$\dot{XP} = r_{pts} - r_{pts_up}$$



Rate laws

$$r_{gly} = k_{gly} G6P$$

$$r_{pdh} = k_{pdh} Prv$$

$$r_{pts} = k_{pts}(PEP(X_0 - XP) - K_{pts} Prv XP)$$

$$r_{pyk} = k_{pyk} PEP f(PEP, \dots)$$

Steady-state conditions

$$r_{gly} = r_{n-pts} + r_{pts_up}$$

$$r_{pdh} = 2(r_{n-pts} + r_{pts_up})$$

$$r_{pyk} = 2r_{n-pts} + r_{pts_up}$$

Steady-state concentrations

Kremling et al.07

$$G6P = \frac{r_{n-pts} + r_{pts_up}}{k_{gly}}$$

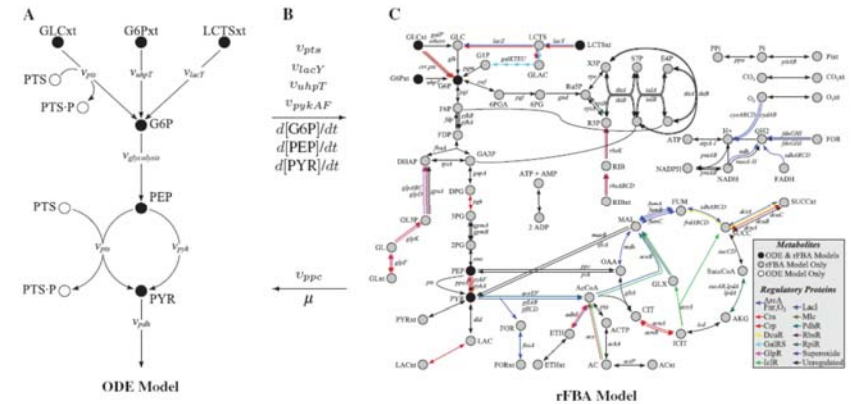
$$P_{TV} = 2 \frac{r_{n-pts} + r_{pts_up}}{k_{pdh}}$$

$$PEP = \frac{2r_{n-pts} + r_{pts_up}}{k_{pykf}}$$

$$XP = k_{pdh} \frac{X_0 - \frac{r_{pts_up}}{k_{pts} PEP}}{k_{pdh} + 2K_{pts} k_{pykf} \frac{r_{n-pts} + r_{pts_up}}{2r_{n-pts} + r_{pts_up}}}$$

Integration of ODE and rFBA model

Covert/Xiao/Chen/Karr'08

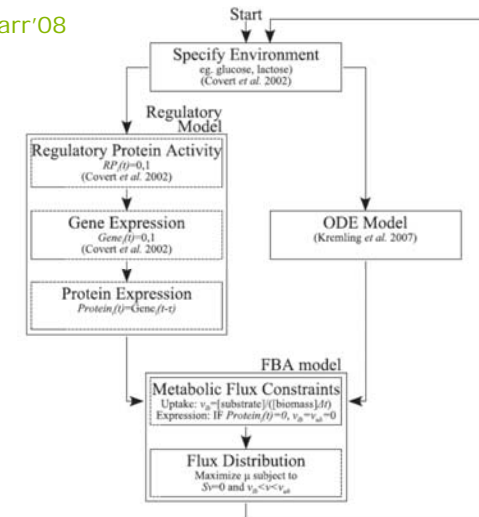


Pass values from one model to the other

- Identify metabolites and fluxes common to both models (black circles in Fig.)
- Variables passed from the ODE model:
 - enzyme fluxes v_{pts} , v_{lacY} , v_{uhpT} , v_{pykAF}
 - changes in metabolite concentrations $d[G6P]/dt$, $d[PEP]/dt$, $d[PYR]/dt$
- Variables passed from the rFBA model:
 - growth flux μ
 - flux through phosphoenolpyruvate carboxylase v_{ppc}

iFBA algorithm

Covert/Xiao/Chen/Karr'08



iFBA algorithm

1. Specify initial environment
2. Calculate regulatory protein activity, gene and protein expression
 - Boolean regulatory model with time delays (Covert et al. 2001)
 - some Boolean regulatory values superseded by ODE model: *Crp*, *galEKMP*, *lacYZ*, *pgk*, *ptsG*, and *pykF*
3. Solve ODEs
 - At each time step, numerically integrate the ODE model using the growth rate and *ppc* flux computed by the FBA model at the previous time step.
 - Calculate ODE rates at the end of the time step to later constrain the FBA linear program.

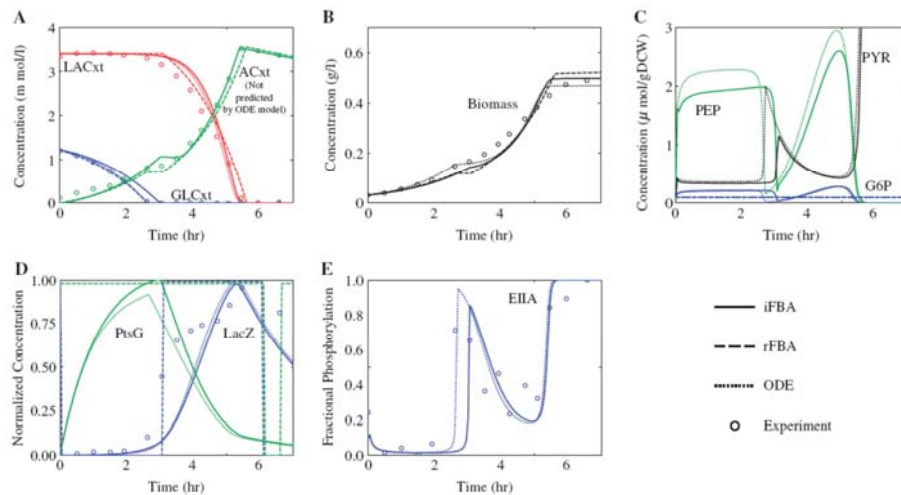
iFBA algorithm

4. Determine metabolic flux constraints and metabolite pooling fluxes
 - Flux constraints:
 - thermodynamic irreversibility
 - environment: available amount of substrate in the culture medium
 - transport: maximum substrate uptake or by-product secretion rate
 - regulation: restrict flux through an enzyme according to the expression of the corresponding protein(s)
 - ODE matching: specification of fluxes by the ODE model
 - Set rhs of FBA linear program to concentrations calculated by ODE model: $d[\text{G6P}]/dt$, $d[\text{PEP}]/dt$, $d[\text{PYR}]/dt$
5. Calculate flux distribution: Maximize biomass production
6. Calculate new environment: Use growth rate and fluxes computed by the FBA model to update biomass and metabolite concentrations

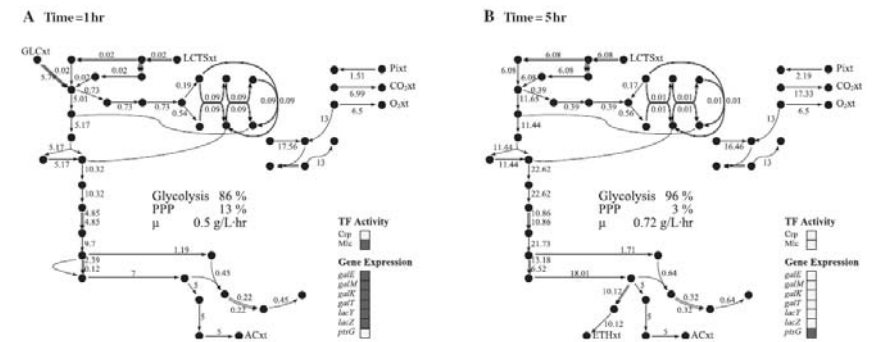
$$[\text{biomass}](t + \Delta t) = \beta [\text{biomass}](t) e^{\mu \Delta t}$$

$$[\text{met}_i](t + \Delta t) = [\text{met}_i] + \frac{v_{ex}}{\mu} [\text{biomass}](t) (1 - e^{\mu \Delta t})$$

Comparing the three approaches



Changes in gene expression and flux distribution

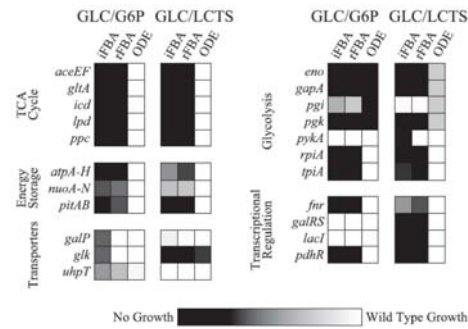


- bacteria are consuming glucose
- significant flux from internal glucose- 6-phosphate through the pentose phosphate pathway

- bacteria are consuming lactose
- flux from glucose-6-phosphate has shifted towards glycolysis
- the lactose-related TFs GalE, GalM, GalK, and GalT are now expressed, while PtsG is suppressed and bacteria secrete ethanol in addition to acetate

Single gene perturbation

- 165 single gene perturbations
- iFBA predicts different phenotypes than ODE for 41 resp. 45 of the mutants on glucose/glucose-6-phosphate resp. glucose/ lactose.
- Corresponding genes can be grouped into 5 classes (see Fig.)
- In most of the 85 cases, iFBA and rFBA predicted the correct phenotype, while ODE failed.
- In 2 cases, iFBA predicted different phenotypes than rFBA (better account of the dynamics of internal metabolites).



Discussion

- Kinetic description in iFBA much more detailed than in rFBA.
- Concentration of internal metabolites not calculable without kinetic model (rFBA uses approximation from external metabolite concentrations or combinations of fluxes).
- iFBA can model enzymes that do not contribute to metabolic growth, but have another important function such as signal transduction ("dead ends" in standard FBA).
- Compared with ODE, iFBA may compute flux distributions for an entire network with only a few parameters -> more predictive power than ODE, both in terms of scope and accuracy.
- ODE models can capture intracellular concentration and short-time dynamics (critical in signal transduction) -> iFBA as an integrating framework.

Possible improvements:

- More accurate objective functions
- Flux variability analysis

References

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 Covert MW, Xiao N, Chen TJ, Karr JR.
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